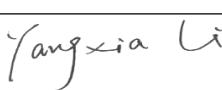


Performance Study Report

<Product name: SARS-CoV-2 Antigen Test Cassette >

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1. Introduction

This test cassette is only used for in vitro qualitative detection of SARS-CoV-2 antigen in human nasopharyngeal, nasal swabs and saliva specimens within 14 days after clinical symptoms. The test cassette is intended for screening of patients suspected for infection with SARS-CoV-2, and as an aid in the diagnosis of the coronavirus disease (COVID-19).

In order to investigate its appearance, physical properties, migration velocity, negative coincidence rate, positive coincidence rate, sensitivity, repeatability(precision), specificity, cross reactivity, interfering substances and limit of detection (LOD), the following tests are carried out according to the requirements of inspection procedure of this product.

2. Information of SARS-CoV-2 Antigen Test Cassette

Three batches of SARS-CoV-2 Antigen Test Cassette were produced by Jiangsu Mole Bioscience Co. Ltd. Batch numbers are M0120101, M0120105, M0120107. The following table shows the components of the test cassette.

Table 1 Main components of the Test Cassette

Components	Usage
SARS-CoV-2-NP antibody	Applied to the nitrocellulose membrane in the test area
Goat anti rabbit Ig G polyclonal antibody	Applied to the nitrocellulose membrane in the control area
SARS-CoV-2 monoclonal antibody	Applied to the conjugate pad
Colloidal gold conjugate	

Packaging Size: 20 Tests/Pack.

3. Quality Inspection Standards

3.1 Appearance

The appearance should be smooth, materials should be firmly attached and the components should be complete.

3.2 Physical Inspection

The width of the test strip is $3.0\pm0.1\text{mm}$, and the volume of the sample extraction solution is not less than 12mL (20 Test).

3.3 Migration Velocity

The migration velocity should not be less than 10mm/min.

3.4 Sensitivity

(1) Sensitivity References Test

Three reference solutions were used for testing, the detection results of three SARS-CoV-2 antigen sensitivity references (S1, S2, S3) require that the reference S1 must be tested as negative, and the references S2 and S3 should be tested as positive.

(2) Clinical Sensitivity

SARS-CoV-2 Antigen Test Cassette and the SARS-CoV-2 detection kit (RT-PCR method) were used to test the same sample to determine its clinical sensitivity.

3.5 Negative Coincidence Rate

Ten SARS-CoV-2 antigen negative references (N1-N10) were tested with the three batches of products, and all test results should be negative.

3.6 Positive Coincidence Rate

Test Five SARS-CoV-2 antigen positive references (P1-P5) were tested with three batches of products, and all the test results should be positive.

3.7 Repeatability (Precision)

The test of samples from three batches of products was repeated 10 times via using the same reference solution (R) and positive solution with different concentrations (P1, P2). The color density of the test line should be uniform and consistent. The test results of negative solution should be all negative and the test results of positive solution should be all positive.

3.8 Specificity

(1) Specificity Testing with Strains

Following strains at specified concentrations were spiked onto swabs (or saliva specimen) and tested according to the Instruction for Use. Read the results at 15 minutes. Results should all be negative.

Table 2 Specificity Testing with Strains

Description	Concentration
Human coronaviruses 229E	2.35×10^6 TCID ₅₀ /ml
Human Coronavirus OC43	2.45×10^6 TCID ₅₀ /ml
Coronavirus NL63	1×10^5 TCID ₅₀ /ml
Influenza A H1N1	3.16×10^5 TCID ₅₀ /ml
Influenza A H3N2	1×10^5 TCID ₅₀ /ml
Influenza B	1×10^5 TCID ₅₀ /ml
Human Rhinovirus 2	2.81×10^4 TCID ₅₀ /ml
Human Rhinovirus 14	1.58×10^6 TCID ₅₀ /ml
Human Rhinovirus 16	8.89×10^6 TCID ₅₀ /ml
Measles virus	1.58×10^4 TCID ₅₀ /ml
Adenovirus 3	1.58×10^4 TCID ₅₀ /ml
Parainfluenza virus 2	1.58×10^6 TCID ₅₀ /ml
Parainfluenza virus 3	1.58×10^7 TCID ₅₀ /ml
Respiratory syncytial virus	8.89×10^4 TCID ₅₀ /ml

(2) Clinical Specificity

Use the SARS-CoV-2 Antigen Test Cassette and the SARS-CoV-2 detection kit (RT-PCR method) to test the same sample to determine its clinical specificity.

3.9 Cross Reactivity

Substances in different concentrations were spiked onto swabs (saliva specimens) and tested according to the Instruction for Use in triplicate. Read results at 15 minutes. Results should be negative.

Table 3 Cross reaction substances

Substances	Concentration
<i>Arcanobacterium bernardiae</i>	1×10^7 CFU/ml
<i>Arcanobacterium haemolyticum</i>	1×10^7 CFU/ml
<i>Candida albicans</i>	1×10^7 CFU/ml
<i>Corynebacterium</i>	1×10^7 CFU/ml
<i>Moraxella catarrhalis</i>	1×10^7 CFU/ml
<i>Neisseria lactamica</i>	1×10^7 CFU/ml
<i>Pseudomonas aeruginosa</i>	1×10^7 CFU/ml
<i>Staphylococcus aureus subsp aureus</i>	1×10^7 CFU/ml
Influenza B (Yamagata and Victoria) virus	3.16×10^6 TCID ₅₀ /ml
Human Rhinovirus 2	2.81×10^4 TCID ₅₀ /ml
Human Rhinovirus 14	1.58×10^6 TCID ₅₀ /ml
Human Rhinovirus 16	8.89×10^6 TCID ₅₀ /ml
Measles virus	1.58×10^4 TCID ₅₀ /ml
Adenovirus 3	1.58×10^4 TCID ₅₀ /ml
Human coronaviruses 229E	2.35×10^6 TCID ₅₀ /ml
<i>Staphylococcus epidermidis</i>	1×10^7 CFU/ml
<i>Streptococcus pneumoniae</i>	1×10^7 CFU/ml
<i>Streptococcus salivarius</i>	1×10^7 CFU/ml
<i>Streptococcus sp. Group F</i>	1×10^7 CFU/ml
Human Coronavirus OC43	2.45×10^6 TCID ₅₀ /ml
Coronavirus NL63	1×10^5 TCID ₅₀ /ml
Influenza A H1N1	3.16×10^5 TCID ₅₀ /ml
Influenza A H3N2	1×10^5 TCID ₅₀ /ml
Parainfluenza virus 2	1.58×10^6 TCID ₅₀ /ml
Parainfluenza virus 3	1.58×10^7 TCID ₅₀ /ml
Respiratory syncytial virus	8.89×10^4 TCID ₅₀ /ml

MERS-coronavirus	10^5 PFU/ml
Epstein-Barr virus	10^5 PFU/ml
human metapneumovirus (hMPV)	2.25×10^5 TCID ₅₀ /ml
Coronavirus HKU1	1×10^5 TCID ₅₀ /ml

3.10 Interfering Substances

We use the substances in the table to perform interference experiments. Repeat the test three times for each batch of the test cassette. The results should be negative.

Table 4 Interfering substances

Analytes	Concentration
Whole Blood	$20\mu\text{l}/\text{ml}$
Mucin	$50\mu\text{g}/\text{ml}$
Budesonide Nasal Spray	$200\mu\text{l}/\text{ml}$
Dexamethasone	$0.8\text{mg}/\text{ml}$
Flunisolide	$6.8\text{ng}/\text{ml}$
Mupirocin	$12\text{mg}/\text{ml}$
Oxymetazoline	$0.6\text{mg}/\text{ml}$
Phenylephrine	$12\text{mg}/\text{ml}$
Sterimar Nasal Spray (Saline)	1:1 (V/V)
Relenza	$282\text{ng}/\text{ml}$
Tamiflu	$1.1\mu\text{g}/\text{ml}$
Tobramycin	$2.43\text{mg}/\text{ml}$
Naso Gel	5% (V/V)
CVS Nasal Spray(Cromolyn)	15% (V/V)

3.11 Determination of Limit of detection (LoD, analytical sensitivity)

To determine the limit of detection, a recombine SARS-CoV-2 nucleocapsid protein was diluted to a series of concentrations, comparing with the SARS-CoV-2 virus (inactivated). The assays were performed according to the Instruction for Use.

4. Performance Evaluation

4.1 Appearance

The appearance of 3 batches test cassette were smooth, and materials were firmly attached and the components were complete.

4.2 Physical Inspection

4.2.1 The width of Test Strip

Measure the width of the test strip with vernier caliper, and the results are shown in the following table:

Table 5 Test results

Batches	M0120101			M0120105			M0120107		
	Repeat	1	2	3	1	2	3	1	2
Strip Width (mm)	3.01	2.98	3.04	2.96	2.96	3.02	3.00	2.99	2.99

Conclusion: The results of experiments showed that the width of the test strip of the SARS-CoV-2 Antigen Test

Clinical sensitivity (positive coincidence rate): $116 / (116+7) \times 100\% = 94.31\%$
 95% confidence interval: [0.8872, 0.9722]

4.5 Negative Coincidence Rate

Test the three batches of kits with negative references (N1-N10), and interpret the results in 15 to 20 minutes.

The results are shown in the following table.

Table 14 The negative coincidence test results

Batches	M0120101			M0120105			M0120107		
	1	2	3	1	2	3	1	2	3
N1	—	—	—	—	—	—	—	—	—
N2	—	—	—	—	—	—	—	—	—
N3	—	—	—	—	—	—	—	—	—
N4	—	—	—	—	—	—	—	—	—
N5	—	—	—	—	—	—	—	—	—
N6	—	—	—	—	—	—	—	—	—
N7	—	—	—	—	—	—	—	—	—
N8	—	—	—	—	—	—	—	—	—
N9	—	—	—	—	—	—	—	—	—
N10	—	—	—	—	—	—	—	—	—

Note: “—” means negative result, “+” means positive result and “±” means critical state.

Conclusion: The results of experiments show that negative coincidence rate of the SARS-CoV-2 Antigen Test Cassette is 100%, which meets the requirements.

4.6 Positive Coincidence Rate

Test the three batches of kits with positive references (P1-P5), and interpret the results in 15 to 20 minutes. The results are shown in the following table:

Table 15 The positive coincidence test results

Batches	M0120101			M0120105			M0120107		
	1	2	3	1	2	3	1	2	3
P1	+	+	+	+	+	+	+	+	+
P2	+	+	+	+	+	+	+	+	+
P3	+	+	+	+	+	+	+	+	+
P4	+	+	+	+	+	+	+	+	+
P5	+	+	+	+	+	+	+	+	+

Note: “—” means negative result, “+” means positive result and “±” means critical state .

Conclusion: The results of experiments show that the positive coincidence rate of the SARS-CoV-2 Antigen Test Cassette is 100%, which meets the requirements.

4.7 Repeatability (Precision)

Repeatedly test the three batches of kits with repeatability reference (R, P1, P2) for 10 times respectively, and interpret the results in 15 to 20 minutes. The results are shown in the following table:

Table 16 The repeatability test results of positive solution(P1)

Batches	M0120101	M0120105	M0120107
Test repeat 1	+	+	+
Test repeat 2	+	+	+
Test repeat 3	+	+	+
Test repeat 4	+	+	+
Test repeat 5	+	+	+
Test repeat 6	+	+	+
Test repeat 7	+	+	+
Test repeat 8	+	+	+
Test repeat 9	+	+	+
Test repeat 10	+	+	+
Color depth	uniform and consistent	uniform and consistent	uniform and consistent

Note: “—” means negative result, “+”means positive result and “±” means critical state.

Table 17 The repeatability test results of positive solution(P2)

Batches	M0120101	M0120105	M0120107
Test repeat 1	+	+	+
Test repeat 2	+	+	+
Test repeat 3	+	+	+
Test repeat 4	+	+	+
Test repeat 5	+	+	+
Test repeat 6	+	+	+
Test repeat 7	+	+	+
Test repeat 8	+	+	+
Test repeat 9	+	+	+
Test repeat 10	+	+	+
Color density	uniform and consistent	uniform and consistent	uniform and consistent

Note: “—” means negative result, “+”means positive result and “±” means critical state.

(2) Clinical Specificity

The compare results between the SARS-CoV-2 Antigen Test Cassette with the commercial RT-PCR detection reagent on the tables below.

Table 20 The nasal swab samples test results

Assessment reagent	Nucleic acid test (PCR)		Total
	Positive (+)	Negative (-)	
Positive (+)	146	1	147
Negative (-)	4	123	127
Total	150	124	274

Clinical specificity (negative coincidence rate): $123 / (1+123) \times 100\% = 99.19\%$

95% Confidence interval: [0.960, 0.999]

Table 21 The nasopharyngeal swab samples test results

Assessment reagent	Nucleic acid test (PCR)		Total
	Positive (+)	Negative (-)	
Positive (+)	116	1	117
Negative (-)	2	120	122
Total	118	121	239

Clinical specificity (negative coincidence rate): $120 / (1+120) \times 100\% = 99.17\%$

95% Confidence interval: [0.955, 0.999]

Table 22 The saliva samples test results

Assessment reagent	Nucleic acid test (PCR)		Total
	Positive (+)	Negative (-)	
Positive (+)	116	0	116
Negative (-)	7	120	127
Total	123	120	243

Clinical specificity (negative coincidence rate): $120 / (0+120) \times 100\% = 100\%$

95% Confidence interval: [0.969, 1.000]

4.9 Cross Reactivity Test Results

Three batches of reagents were tested three times with potential different concentrations of the cross substances shown in the following table respectively, and the results were consistent, and interpret the results in 15 to 20 minutes. The results are shown in the following table.

Table 23 The Cross-reactivity evaluation results

Substances	Concentration	Batch No					
		M0120101	M0120105	M0120107			
<i>Arcanobacterium</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Candida albicans</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Corynebacterium</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Escherichia coli.</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Moraxella catarrhalis</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Neisseria lactamica</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Nessereria subflava</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Pseudomonas aeruginosa</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Staphylococcus aureus subspauseus</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Staphylococcus epidermidis</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Streptococcus pneumoniae</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Streptococcus pygenes</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Streptococcus salivarius</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
<i>Streptococcus sp group F</i>	1x10 ⁸ org/ml	—	—	—	—	—	—
Human Coronavirus OC43	2.45 x 10 ⁶ LD ₅₀ /ml	—	—	—	—	—	—
Coronavirus NL63	1 x 10 ^{5.07} U/ml	—	—	—	—	—	—
Influenza A H1N1	3.16 x 10 ⁵ TCID ₅₀ /ml	—	—	—	—	—	—
Influenza A H3N2	1 x 10 ⁵ TCID ₅₀ /ml	—	—	—	—	—	—
Influenza B (Yamagata and Victoria) virus	3.16 x 10 ⁶ TCID ₅₀ /ml	—	—	—	—	—	—
Human Rhinovirus 2	2.81 x 10 ⁴ TCID ₅₀ /ml	—	—	—	—	—	—
Human Rhinovirus 14	1.58 x 10 ⁶ TCID ₅₀ /ml	—	—	—	—	—	—
Human Rhinovirus 16	8.89 x 10 ⁶ TCID ₅₀ /ml	—	—	—	—	—	—
Measles virus	1.58x 10 ⁴ TCID ₅₀ /ml	—	—	—	—	—	—
Adenovirus 3	1.58 x 10 ⁴ TCID ₅₀ /ml	—	—	—	—	—	—
Parainfluenza virus 2	1.58 x 10 ⁷ TCID ₅₀ /ml	—	—	—	—	—	—
Parainfluenza virus 3	1.58 x 10 ⁸ TCID ₅₀ /ml	—	—	—	—	—	—
Respiratory syncytial virus	8.89 x 10 ⁴ TCID ₅₀ /ml	—	—	—	—	—	—
MERS-coronavirus	10 ⁵ PFU/ml	—	—	—	—	—	—
EB virus	10 ⁵ PFU/ml	—	—	—	—	—	—

Note: “—” means negative result, “+”means positive result and “±” means critical state.

4.10 Interfering Substances Results

We use the substances in the table to perform interference experiments. Repeat the test three times for each batch of the test cassette. The results should be negative.

Table 26-2 The test results of SARS-CoV-2 nucleocapsid protein

Positive Batches	100ng/ml	10ng/ml	1ng/ml	500pg/ml	200pg/ml	100pg/ml	50pg/ml
M0120105	Test 1	+	+	+	+	+	-
	Test 2	+	+	+	+	+	-
	Test 3	+	+	+	+	+	-
	Test 4	+	+	+	+	+	-
	Test 5	+	+	+	+	+	-
	Test 6	+	+	+	+	+	-
	Test 7	+	+	+	+	+	-
	Test 8	+	+	+	+	+	-
	Test 9	+	+	+	+	+	-
	Test 10	+	+	+	+	+	-
	Test 11	+	+	+	+	+	-
	Test 12	+	+	+	+	+	-
	Test 13	+	+	+	+	+	-
	Test 14	+	+	+	+	+	-
	Test 15	+	+	+	+	+	-
	Test 16	+	+	+	+	+	-
	Test 17	+	+	+	+	+	-
	Test 18	+	+	+	+	+	-
	Test 19	+	+	+	+	+	-
	Test 20	+	+	+	+	+	-
	Test 21	+	+	+	+	+	-
	Test 22	+	+	+	+	+	-
	Test 23	+	+	+	+	+	-
	Test 24	+	+	+	+	+	-
	Test 25	+	+	+	+	+	-
	Test 26	+	+	+	+	+	-
	Test 27	+	+	+	+	+	-
	Test 28	+	+	+	+	+	-
	Test 29	+	+	+	+	+	-
	Test 30	+	+	+	+	+	-

Table 27-3 The test results of SARS-CoV-2 nucleocapsid protein

Batches \ Positive	100ng/ml	10ng/ml	1ng/ml	500pg/ml	200pg/ml	100pg/ml	50pg/ml
M0120107	Test 1	+	+	+	+	+	-
	Test 2	+	+	+	+	+	-
	Test 3	+	+	+	+	+	-
	Test 4	+	+	+	+	+	-
	Test 5	+	+	+	+	+	-
	Test 6	+	+	+	+	+	-
	Test 7	+	+	+	+	+	-
	Test 8	+	+	+	+	+	-
	Test 9	+	+	+	+	+	-
	Test 10	+	+	+	+	+	-
	Test 11	+	+	+	+	+	-
	Test 12	+	+	+	+	+	-
	Test 13	+	+	+	+	+	-
	Test 14	+	+	+	+	+	-
	Test 15	+	+	+	+	+	-
	Test 16	+	+	+	+	+	-
	Test 17	+	+	+	+	+	-
	Test 18	+	+	+	+	+	-
	Test 19	+	+	+	+	+	-
	Test 20	+	+	+	+	+	-
	Test 21	+	+	+	+	+	-
	Test 22	+	+	+	+	+	-
	Test 23	+	+	+	+	+	-
	Test 24	+	+	+	+	+	-
	Test 25	+	+	+	+	+	-
	Test 26	+	+	+	+	+	-
	Test 27	+	+	+	+	+	-
	Test 28	+	+	+	+	+	-
	Test 29	+	+	+	+	+	-
	Test 30	+	+	+	+	+	-

Note: "+" mean positive result, "-" mean negative result.

Table 28 The test results of SARS-CoV-2 virus (inactivated)

Positive Batches		100 TCID ₅₀ /ml	50 TCID ₅₀ /ml	25 TCID ₅₀ /ml
M0120101	Test 1	+	+	-
	Test 2	+	+	-
	Test 3	+	+	-
	Test 4	+	+	-
	Test 5	+	+	-
	Test 6	+	+	-
	Test 7	+	+	-
	Test 8	+	+	-
	Test 9	+	+	-
	Test 10	+	+	-
M0120105	Test 1	+	+	-
	Test 2	+	+	-
	Test 3	+	+	-
	Test 4	+	+	-
	Test 5	+	+	-
	Test 6	+	+	-
	Test 7	+	+	-
	Test 8	+	+	-
	Test 9	+	+	-
	Test 10	+	+	-
M0120107	Test 1	+	+	-
	Test 2	+	+	-
	Test 3	+	+	-
	Test 4	+	+	-
	Test 5	+	+	-
	Test 6	+	+	-
	Test 7	+	+	-
	Test 8	+	+	-
	Test 9	+	+	-
	Test 10	+	+	-

Note: "+" mean positive result, "-" mean negative result.

Conclusion: The minimal limit of detection of SARS-CoV-2 Antigen Test Cassette is 100pg/ml for recombine SARS-CoV-2 nucleocapsid protein. Minimal limit of detection of SARS-CoV-2 Antigen Test Cassette is 50 TCID₅₀/ml for SARS-CoV-2 virus.

5. Conclusion

The physical properties, migration velocity, negative coincidence rate, positive coincidence rate, sensitivity, repeatability(precision), specificity, cross reactivity, interfering substances and limit of detection (LOD) of the SARS-CoV-2 Antigen Test Cassette meet the product quality inspection standards and are all qualified.